Introduction: Peak and trough serum concentrations of vancomycin were determined in term newborn infants with confirmed or suspected *Staphylococcus* sp sepsis by high performance liquid chromatography and fluorescence polarization immunoassay.

Objective: To statistically compare the results of the high performance liquid chromatography and fluorescence polarization immunoassay techniques for measuring serum vancomycin concentrations.

Methods: Eighteen peak and 20 trough serum samples were assayed for vancomycin concentrations using high performance liquid chromatography and fluorescence polarization immunoassay from October 1995 to October 1997.

Results: The linear correlation coefficients for high performance liquid chromatography and fluorescence polarization immunoassay were 0.27 (peak, \( P = 0.110 \)) and 0.26 (trough, \( P = 0.1045 \)) respectively, which were not statistically significant.

Conclusion: There was wide variation in serum vancomycin concentrations determined by high performance liquid chromatography as compared with those determined by fluorescence polarization immunoassay. There was no recognizable pattern in the variability; in an apparently random fashion, the high performance liquid chromatography measurement was sometimes substantially higher than the fluorescence polarization immunoassay measurement, and at other times it was substantially lower.


The rising incidence of nosocomial methicillin-resistant *Staphylococcal* sp infections in intensive care units has resulted in the widespread use of vancomycin, which is a potentially ototoxic and nephrotoxic when serum concentrations are not kept within narrow therapeutic ranges (peak concentrations between 20-40 µg/mL and trough between 5-10 µg/mL)\(^{1-3}\). This toxicity is of particular concern in newborns and patients with kidney diseases, liver diseases, and neoplasias, since they normally receive other nephrotoxic drugs that may potentiate vancomycin effects. In these groups of patients, vancomycin monitoring is routinely performed to minimize risks\(^4-6\).

Two techniques were compared in the present study: fluorescence polarization immunoassay (FPIA) (TDX / Abbott), and high performance liquid chromatography (HPLC). FPIA is more commonly used in clinical laboratories due to the ease of performance, tiny serum volumes required, rapidity, and lower cost in comparison to HPLC. However, there are some reports on the existence of an inactive vancomycin metabolite that is not distinguished by FPIA, while it is separately evaluated by HPLC. This metabolite has been found in patients undergoing peritoneal dialysis\(^7-9\). Its serum concentration, as
well as its depuration, depends mainly on the half-life of vancomycin in the circulation and on blood pH. Consequently, although FPIA employs monoclonal antibodies directed to vancomycin, this technique could possibly overestimate vancomycin concentrations in specific groups of patients.

We compared peak and trough serum vancomycin concentrations as measured by FPIA and HPLC and evaluated their association with regard to clinical efficacy and toxicity.

PATIENTS AND METHODS

Twenty term newborn infants (gestational age 37 to 42 weeks) receiving vancomycin due to a confirmed or presumed *Staphylococcal* sp sepsis, were enrolled in the study from October 1995 to October 1997.

Diagnostic criteria for neonatal sepsis were based on clinical evaluation, presence of hypothermia, fever, respiratory distress (tachypnea, grunting, apnea), cyanosis, tachycardia, poor feeding, gastric residue, and abdominal distension. Newborns presenting with renal failure or receiving other potentially nephrotoxic drugs such as amikacin, amphotericin B, furosemide, or indomethacin were excluded. Vancomycin doses were determined according to the recommendations of Young and Mangun, as well as according to gestational age. Vancomycin was diluted in a 5% dextrose water solution, and administration occurred over a 60-minute period, controlled by an infusion pump. Seventy-two hours after the onset of treatment, two 0.5 mL blood samples and two 1.0 mL blood samples were used to determine vancomycin concentration by FPIA and HPLC, respectively. Peak samples were obtained 1 hour after antibiotic infusion, and trough samples were obtained 1 hour before the next infusion. FPIA was performed according to the manufacturer’s instructions, and HPLC was performed using a Shimadzu 6A model chromatographic apparatus with a 230 nm UV detector, CLC-ODS 150 X 6 mm reverse phase column (0.05 M, pH 4.6 phosphate-buffer mobile phase, methanol, acetonitrile 80:15:5) with a 0.8 mL/min flux.

RESULTS

The studied population consists of 20 term newborn infants receiving vancomycin. Table 1 shows the descriptive statistics (mean ± SD, median, minimum, and maximum) of the parameters measured.

Table 2 lists the descriptive statistics (mean ± SD, median, minimum, and maximum) of the doses used and the gestational ages of the infants.

Table 3 shows the analysis of concordance of the finding of normality—the adequacy of serum concentrations achieved—for peak serum vancomycin concentrations as determined by the FPIA method compared with the finding of normality by the HPLC method. There was no statistically significant concordance (as determined by the McNemar test, \( P = 0.4531 \)) regarding normality of peak concentrations of serum vancomycin between the FPIA and HPLC methods.

Table 4 shows the analysis of concordance of the finding of normality for trough serum vancomycin concentrations as determined by the FPIA method compared with the finding of normality.
DISCUSSION

There are a few prospective studies on monitoring of vancomycin in newborn infants, and in most cases they used the FPIA method. It is accepted that initial doses of vancomycin should be calculated on the basis of postconceptual age and body weight. Dose corrections are afterwards made by monitoring of serum concentrations of vancomycin. This practice decreases drug toxicity and also the use of insufficient dosages, thus favoring newborn infant’s successful treatment. However, measured vancomycin concentrations may be dramatically affected by the method used; FPIA is less time consuming but has the disadvantage of overestimating drug concentrations due to the presence of an inactive metabolite that is detected together with the intact drug, even though a monoclonal antibody is employed in this commercial kit. This problem has not yet been reported concerning HPLC, which is unfortunately a methodology that is not feasible in a clinical laboratory. In our study, we were unable to use HPLC clinically, so dose modifications were based on FPIA results.

When normality (adequate serum concentrations of vancomycin) was correlated for FPIA vs. HPLC, we did not find statistically significant correlations for either peak or trough concentrations.

We also did not find a statistically significant linear correlation of the FPIA vs HPLC methods for either peak or trough serum concentrations of vancomycin.

CONCLUSION

Monitoring of serum concentrations of vancomycin is mandatory for monitoring vancomycin therapy in
both term and premature newborn infants, since in these patients there are rapid and intermittent changes in body water composition and renal clearance that interfere with drug kinetics, distribution, metabolism, and depuration. When interpreting the results of serum vancomycin concentration assays, neonatologists should consider the advantages and disadvantages of each method; FPIA possibly overestimates concentrations in certain cases, and HPLC is not a feasible method in clinical laboratories. However, whatever the method chosen, it is important to highlight that the present study showed a lack of linear correlation between FPIA and HPLC for both peak and trough vancomycin concentrations.

REFERENCES


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