GROWTH RETARDANTS ON DEVELOPMENT AND ORNAMENTAL QUALITY OF POTTED ‘LILLIPUT’ Zinnia elegans JACQ.

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ABSTRACT: Zinnias have good potential to be used as flowering, potted plants, being a quick source of novelty for the floriculture industry with the aid of growth retardants. This study evaluated the effect of growth retardants on development and production of short, compact and attractive plants of potted ‘Lilliput’ Zinnia elegans, a highly ornamental zinnia with low cost seeds. Trials were set up in randomized blocks, with ten treatments (control and three treatments of each retardant: daminozide, paclobutrazol and chlormequat) and four replications (two pots per experimental unit, with one plant per 0.6-L pot). Paclobutrazol (0.5, 0.75 and 1.0 mg a.i. per pot) and chlormequat (1.0, 2.0 and 3.0 g L⁻¹) were applied as a single drench (40 mL per pot), and daminozide (2.5, 3.75 and 5.0 g L⁻¹) as a single foliar spray to runoff (10 mL per pot), at apical flower bud stage. Daminozide (2.5 and 3.75 g L⁻¹), paclobutrazol (0.5, 0.75 and 1.0 mg a.i. per pot) and chlormequat at 1.0 g L⁻¹ significantly reduced plant height and side branches length, without affecting flower diameter, delaying production cycle and causing phytotoxicity symptoms. However, plants were not short and compact enough to meet market quality demand. Chlormequat (2.0 and 3.0 g L⁻¹) caused phytotoxicity symptoms and daminozide (5.0 g L⁻¹) delayed production cycle.

Key words: floriculture, daminozide, paclobutrazol, chlormequat, flowering

INTRODUCTION

Zinnias are well known garden plants and cut flowers in Brazil, and can also be used as flowering potted plant. They have attractive capitula with a wide variety of colors, shape and size, long bloom period, drought tolerance, easy propagation, fast growth, minimal labor requirements, rusticity, and are a quick source of novelty for the floriculture industry with the aid of growth retardants (Pinto, 2003). Zinnia elegans ‘Lilliput’ is a dwarf, short garden plant (31.0 to 46.0 cm in height) (Metcalf & Sharma,
It blooms a wide variety of brightly, bi-colored ray florets and a highly double-flowered capitula type, which increase its ornamental value compared to single and semi-double zinnia type capitula (Miyajima & Nakayama, 1994). Studies on production of zinnias as potted plants showed that ‘Lilliput’ was not short and compact enough to produce high-quality, marketable plants without the aid of growth retardants (Pinto, 2003), commonly used to produce short and attractive compact floriferous plants (Andersen & Andersen, 2000).

Studies on the effect of growth retardants on *Zinnia elegans* cultivars are only a few (Armitage et al., 1981; Banko & Stefani, 1988; Cox & Keever, 1988; Latimer, 1991; Kim et al., 1992; Chen et al., 1993), none of them done with ‘Lilliput’. Effects of growth retardants on *Zinnia elegans* have not been previously studied in Brazil. Differences in responsiveness to growth retardants may be found among cultivars and can depend on environmental and cultural growing conditions (Grzesiski, 1989). The objective of this study was to evaluate the effects of growth retardants on development and ornamental characteristics of potted zinnia ‘Lilliput’, aiming to produce short and attractive compact plants.

**MATERIAL AND METHODS**

Plants were grown in the Fall of 2001, in a polyethylene-covered greenhouse, in Jaboticabal, SP, Brazil (21°15’22” S, 48°15’18” W; altitude 610 m). Seeds of ‘Lilliput’ *Zinnia elegans* were sown on polystyrene plug trays (model CM 128-62), filled with commercial media and transplanted to 0.6-L black plastic pots (9.4 cm deep; 13.0 cm wide), when second nodal leaves were fully expanded. Each pot was filled with commercial growing media: composted pine and eucalyptus bark, composted tree fern fiber dust, washed coconut dust, sterilized rice media: composted pine and eucalyptus bark, composted

Main stem height (from main stem base to inflorescence receptacle insertion), internode length, number of nodes, side branches length and total leaf area were measured at the end of production cycle (six to seven open inflorescences per plant). Total leaf area was estimated by taking a sample of 20 leaves from each experimental unit, measured with a Digital Image Analysis System, Delta T Devices. Total leaf area was determined by proportion, using the dry mass value of measured leaves.

Production cycle was established considering the number of days from sowing to six or seven opened inflorescences. Opened inflorescence was considered when ray florets petals were fully expanded, and tubular florets opened. To evaluate plant quality, inflorescence diameter, florets petals were fully expanded, and tubular florets opened. To evaluate plant quality, inflorescence diameter, plant height and plant spread diameter (major width from one tip to the other of foliage canopy) were measured.

Data were subjected to analysis of variance (ANOVA) and to Tukey’s multiple range test. Regression analysis was determined to establish the concentration response curves of growth retardants (control data included in regression analysis). Polynomial regression analyses were performed for all evaluated parameters.

**RESULTS AND DISCUSSION**

Chlormequat drench at 2.0 and 3.0 g L\(^{-1}\) caused a slightly visible wilt (Figure 1-A), 15 h after application. Pale green aqueous lesions were observed on leaf lamina margins 24 h after application (Figure 1-B), followed by foliar chlorosis on leaf margins and center, which persisted during production period (2.0 g L\(^{-1}\)), making plants unsuitable for marketing. Plants treated with chlormequat at 3.0 g L\(^{-1}\) developed severe foliar chlorosis and necrosis (Figure 1-C), leading to plants death. ‘Lilliput’ was sensitive to chlormequat concentrations in the range of 2.0 to 3.0 g L\(^{-1}\). However, Cathey (1975) reported that concentration ranging on 0.5 to 3.0 g L\(^{-1}\) were


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inactive to ‘Yellow Zenith’ Z. elegans. No visual, phytotoxic symptoms were observed on plants treated with 1.0 g L⁻¹ of chlormequat other than thicker roots compared to control (Figure 1-D and Figure 1-E).

Plants treated with paclobutrazol showed darker green leaves in comparison to control, daminozide and chlormequat (1.0 g L⁻¹) treated plants. This is a common effect for paclobutrazol-treated plants as a result of either increasing chlorophyll biosynthesis and/or reduction of leaf expansion accompanied by normal rates of chlorophyll biosynthesis (Davis et al., 1988; Halmann, 1990). There were no apparent effects of daminozide, paclobutrazol and chlormequat (1.0 g L⁻¹) on inflorescence color and shape.

At the end of production cycle, treated plants were shorter than control plants. Main stem height suppression was related to shorter internode elongation and not to a decrease in node number (Table 1). Internode length and main stem height responded quadratically to daminozide treatment, but showed a negative linear re-

Table 1 - Effects of growth retardants on main stem internode length, height and number of nodes, side branches length, plant height and spread diameter of ‘Lilliput’ plants, at the end of the production cycle.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Main stem</th>
<th></th>
<th>Side branches</th>
<th>Plant</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Internode length</td>
<td>Height</td>
<td>No. of nodes</td>
<td>Length</td>
</tr>
<tr>
<td>Control</td>
<td>6.51 a</td>
<td>51.63 a</td>
<td>7.25 a</td>
<td>30.56 a</td>
</tr>
<tr>
<td>2.50 g L⁻¹ daminozide</td>
<td>4.40 b</td>
<td>34.70 b</td>
<td>7.13 a</td>
<td>18.51 c</td>
</tr>
<tr>
<td>3.75 g L⁻¹ daminozide</td>
<td>4.63 b</td>
<td>38.76 b</td>
<td>7.38 a</td>
<td>21.22 bc</td>
</tr>
<tr>
<td>5.00 g L⁻¹ daminozide</td>
<td>4.96 b</td>
<td>38.46 b</td>
<td>6.88 a</td>
<td>19.53 bc</td>
</tr>
<tr>
<td>0.50 mg a.i. paclobutrazol</td>
<td>5.00 b</td>
<td>41.74 b</td>
<td>7.38 a</td>
<td>26.95 ab</td>
</tr>
<tr>
<td>0.75 mg a.i. paclobutrazol</td>
<td>4.63 b</td>
<td>37.59 b</td>
<td>7.25 a</td>
<td>22.46 bc</td>
</tr>
<tr>
<td>1.00 mg a.i. paclobutrazol</td>
<td>4.63 b</td>
<td>36.99 b</td>
<td>7.38 a</td>
<td>19.50 bc</td>
</tr>
<tr>
<td>1.00 g L⁻¹ chlormequat</td>
<td>4.82 b</td>
<td>36.45 b</td>
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<td>15.27 c</td>
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<tr>
<td>Mean</td>
<td>4.95</td>
<td>39.54</td>
<td>7.16</td>
<td>21.75</td>
</tr>
</tbody>
</table>

F test treatment: 8.35** 10.61** 1.47ns 9.14** 10.29** 1.73ns
HSD (Tukey’s test): 1.09 7.71 1.07 7.69 12.58 6.76

Mean separation within columns by Tukey’s multiple range test, 5% level; HSD = honestly significant difference; “nonsignificant at (P > 0.05); “difference at (P < 0.01).

Figure 1 - A) ‘Lilliput’ control and chlormequat treated plants (1.0; 2.0 and 3.0 g L⁻¹); B) Pale green aqueous lesions on ‘Lilliput’ leaf lamina margin treated with chlormequat (2.0 g L⁻¹); C) Severe leaf burning and necrosis on ‘Lilliput’ leaf lamina treated with chlormequat (3.0 g L⁻¹); D) ‘Lilliput’ control plants root system; E) Root system with thicker roots of ‘Lilliput’ treated plant with chlormequat (1.0 g L⁻¹).
sponse to increasing concentrations of paclobutrazol (Figure 2). Minimum internode length of 4.40 cm for 3.24 g L$^{-1}$ of daminozide and minimum main stem height of 35.78 cm for 3.35 g L$^{-1}$ of daminozide can be predicted from the models. Regression analysis of node number and daminozide and paclobutrazol treatments were not significant (data not shown).

Growth retardants, except paclobutrazol at 0.5 mg a.i. per pot, reduced side branches length and plant height (Table 1). Side branches length and plant height responded quadratically to daminozide treatments but showed a negative linear response to increasing concentration of paclobutrazol (Figure 2). Minimum side branches length of 18.98 cm for 3.71 g L$^{-1}$ of daminozide, and minimum plant height of 51.50 cm for 3.53 g L$^{-1}$ of daminozide, can be predicted from the models (Figure 2).

Banko & Stefani (1988) found that daminozide spray (5.0 g L$^{-1}$) reduced internode length and plant height of ‘Yellow Marvel’ Z. elegans. Daminozide sprays also presented similar results on ‘Lilliput’, but at lower concentrations (2.5 and 3.75 g L$^{-1}$). Banko & Stefani (1988) reported also that paclobutrazol drench (0.15 mg a.i. per pot) had no effect on zinnia growth suppression, requiring higher rates of this compound. In this study, paclobutrazol drench at higher rates (0.5, 0.75 and 1.0 mg a.i. per pot) resulted in shorter plants with shorter internodes. Cox & Keeever (1988) also observed height reduc-

![Figure 2](image_url)

**Figure 2** - Regression models of daminozide and paclobutrazol effects on internode length (A-B), main stem height (C-D), side branches length (E-F) and plant height (G-H) of ‘Lilliput’, at the end of production cycle. Vertical bars represent the standart error of mean, n = 4; *significant difference at ($P < 0.05$); **significant difference at ($P < 0.01$).
tions of ‘Scarlet Ruffles’ *Z. elegans* with paclobutrazol drenches in the range of 0.5 and 1.0 mg a.i. per pot.

One application of paclobutrazol at higher concentrations should be tested, once the polynomial regression analysis for main stem height, internode length, plant height and side branches length showed a negative linear response to the studied concentrations (Figure 2). Plant spread diameter was not affected by growth retardants (Table 1). Plants were too spread (Figure 3). Christensen & Friis (1987) reported that a spread diameter of 20.0 cm would be satisfactory for many flowering potted plants.

Number of leaves and total leaf area were not affected by retardants application, and the regression analysis for these parameters and growth retardants concentrations were not significant (data not shown). Number of leaves of *Z. elegans* is related to plant’s node number (usually, a maximum of two leaves develop at each node). Therefore, when retardants suppress internode elongation without changing the node number, no effect is observed in the number of plant leaves. Although inhibition of gibberellin biosynthesis by growth retardants may rapidly reduce stem elongation, leaf initiation may not be affected if retardants reduce subapical meristematic activity without concomitant disruption of apical meristematic function (Sachs & Hackett, 1972).

Growth retardants altered ‘Lilliput’ dry mass distribution pattern without changing plant total dry mass (Table 2). Plants treated with daminozide (3.75 g L⁻¹),

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Green leaves</th>
<th>Senescent and dead leaves*</th>
<th>Main stem</th>
<th>Side branches</th>
<th>Inflorescences</th>
<th>Floral buds*</th>
<th>Roots</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>22.4 b</td>
<td>1.7 a</td>
<td>20.1 a</td>
<td>29.2 a</td>
<td>21.3 a</td>
<td>1.9 a</td>
<td>5.0 a</td>
<td>5.84 a</td>
</tr>
<tr>
<td>2.50 g L⁻¹ daminozide</td>
<td>24.9 ab</td>
<td>1.8 a</td>
<td>16.8 b</td>
<td>23.6 bcd</td>
<td>24.7 a</td>
<td>2.0 a</td>
<td>6.3 a</td>
<td>5.38 a</td>
</tr>
<tr>
<td>3.75 g L⁻¹ daminozide</td>
<td>27.6 a</td>
<td>1.7 a</td>
<td>18.0 ab</td>
<td>23.7 bcd</td>
<td>22.6 a</td>
<td>1.9 a</td>
<td>5.9 a</td>
<td>5.70 a</td>
</tr>
<tr>
<td>5.00 g L⁻¹ daminozide</td>
<td>25.8 ab</td>
<td>1.7 a</td>
<td>17.3 ab</td>
<td>22.7 bcd</td>
<td>25.4 a</td>
<td>2.0 a</td>
<td>6.0 a</td>
<td>5.70 a</td>
</tr>
<tr>
<td>0.50 mg a.i. paclobutrazol</td>
<td>25.0 ab</td>
<td>1.6 a</td>
<td>17.5 ab</td>
<td>27.5 ab</td>
<td>22.0 a</td>
<td>1.9 a</td>
<td>6.3 a</td>
<td>5.50 a</td>
</tr>
<tr>
<td>0.75 mg a.i. paclobutrazol</td>
<td>26.2 ab</td>
<td>1.6 a</td>
<td>17.5 ab</td>
<td>24.8 abc</td>
<td>23.5 a</td>
<td>1.9 a</td>
<td>5.8 a</td>
<td>4.93 a</td>
</tr>
<tr>
<td>1.00 mg a.i. paclobutrazol</td>
<td>27.0 a</td>
<td>1.7 a</td>
<td>18.4 ab</td>
<td>21.5 cd</td>
<td>24.8 a</td>
<td>1.9 a</td>
<td>6.1 a</td>
<td>4.79 a</td>
</tr>
<tr>
<td>1.00 g L⁻¹ chlormequat</td>
<td>27.4 a</td>
<td>1.6 a</td>
<td>17.4 ab</td>
<td>18.8 d</td>
<td>27.2 a</td>
<td>2.1 a</td>
<td>6.3 a</td>
<td>4.84 a</td>
</tr>
<tr>
<td>Mean</td>
<td>25.8 a</td>
<td>1.7</td>
<td>17.9</td>
<td>24.0</td>
<td>23.9 a</td>
<td>1.9 a</td>
<td>6.0 a</td>
<td>5.34 a</td>
</tr>
</tbody>
</table>

F test treatment | 4.17** | 1.23** | 2.19** | 8.36** | 1.24** | 1.07** | 0.52** | 1.78** |
HSD (Tukey’s test) | 3.99 | 0.21 | 3.26 | 5.39 | 8.36 | 0.33 | 2.81 | 1.50 |
C.V (%) | 6.54 | 5.20 | 7.70 | 9.47 | 14.73 | 7.09 | 19.83 | 11.87 |

Mean separation within columns by Tukey’s multiple range test, 5% level; HSD = honestly significant difference; C.V = coefficient of variation; “nonsignificant at (P > 0.05); ”difference at (P < 0.01); 'data converted to ln(x+5.0), for analysis and presentation.

Figure 3 - Effects of paclobutrazol (A: 1 - control; 2 - 0.5 mg a.i. per pot; 3 - 0.75 mg a.i. per pot; 4 - 1.0 mg a.i. per pot), daminozide (B: 1- control; 2 - 2.5 g L⁻¹; 3 - 3.75 g L⁻¹; 4 - 5.0 g L⁻¹) and chlormequat (C: 1- control; 2 - 1.0 g L⁻¹) on plant height of *Zinnia elegans* ‘Lilliput’, at the end of production cycle.
paclobutrazol (1.0 mg a.i. per pot) and chlormequat (1.0 g L\(^{-1}\)) had greater green leaves dry mass and smaller side branches dry mass, compared to control plants. Only 2.5 g L\(^{-1}\) daminozide-treated plants showed a reduction in main stem dry mass.

There was linear increase in green leaves dry mass and a linear decrease in side branches dry mass, with increasing daminozide and paclobutrazol concentrations. Daminozide also conditioned linear decrease in main stem dry mass and a linear increase in inflorescences dry mass. Increasing paclobutrazol concentration provided a linear increase in root dry mass (Figure 4).

Daminozide and paclobutrazol strongly reduced sink activity in side branches, leading to assimilate retention on leaves. Growth retardants could reduce sink activity in stems while increases in roots and inhibit assimilated \(^1\)\(^3\)C translocation from leaves to others zinnia organs (Kim & Suzuki, 1989). Hence, branches translocation is more restricted compared to roots. Roots growth is less affected, or could also increase, by retardants application (Latimer, 1991). Some authors suggested that gibberellin promotes the increase of sink activity or rates of phloem unloading in the stems, but does not affect assimilate production or phloem loading at the source (Lovell, 1971; Mulligan & Patrick, 1979).

**Figure 4 - Regression models of daminozide and paclobutrazol effects on green leaves (A-B), side branches (C-D), main stem (E), root (F) and inflorescence (G) dry mass of ‘Lilliput’, at the end of production cycle. Vertical bars represent the standart error of mean, n = 4; *significant difference at (\(P < 0.05\)); **significant difference at (\(P < 0.01\)).**
Plants treated with daminozide (3.75 g L\(^{-1}\)), paclobutrazol (1.0 mg a.i. per pot) and chlormequat (1.0 g L\(^{-1}\)) had higher leaf mass ratio compared to control (Table 3). There was linear increase in leaf mass ratio with increasing paclobutrazol and daminozide concentrations (Figure 5), that is, retardants might have induced an assimilate retention on the leaves.

Daminozide (5.0 g L\(^{-1}\)), paclobutrazol (1.0 mg a.i. per pot) and chlormequat (1.0 g L\(^{-1}\)) increased foliage and inflorescence harvest index (Table 3), with a positive, linear response to increasing concentrations of daminozide and paclobutrazol (Figure 5). Although inflorescences harvest index did not differ among control and retardant treatments (Table 3), a linear increase was observed in inflorescences harvest index with increasing daminozide concentrations (Figure 5). This seems to indicate that paclobutrazol induces the conversion of photosynthesis products only in foliage, once no significant tendency was observed between inflorescences harvest index and paclobutrazol concentrations (data not shown). At any rate, daminozide leads to a conversion of photosynthesis products to foliage and inflorescences, once a linear increase in inflorescences harvest index with increasing daminozide concentrations was observed (Table 3). Thus, daminozide tends to promote the translocation of photosynthates also to the inflorescences, which will contribute to improve plant quality, since for potted plants, the harvest products of economic importance are foliage and inflorescences.

‘Lilliput’ inflorescence diameter was not affected by growth retardants (Table 3), and regression analysis for inflorescence diameter and retardants concentrations were not significant (data not shown). However, Armitage et al. (1981) observed reduction on inflorescence diameter of Z. elegans cultivars as the number of daminozide (5.0 g L\(^{-1}\)) applications increased. Chen et al. (1993) recorded that a single foliar application of paclobutrazol (0.25 or 0.75 g L\(^{-1}\)) reduced inflorescence diameter of toasted ‘Sunrise Yellow’ Z. elegans. The number of application (1, 2, 3 and 4 times in the case of daminozide), differences in environmental conditions, cultivars sensibility to retardants, and method of application (spray or drench) may explain these contrastant results.

Daminozide (5.0 g L\(^{-1}\)) delayed production cycle (Table 3), with a positive linear response to increasing concentrations of daminozide (Figure 5). Daminozide delayed inflorescence development (data not shown). Armitage et al. (1981) and Latimer (1991) also found that daminozide at 5.0 g L\(^{-1}\) delayed flowering time in other Z. elegans cultivars. Flowering delay after daminozide application could derive from a blockage of gibberellin synthesis or from a wholly or partly restriction in action of one or more active endogenous gibberellins, which may be flowering promoters. Growth retardants action can be relatively specific for each gibberellin (Koranski et al., 1979). Thus, growth retardant could decrease the synthesis of gibberellin, that promote main stem and side branches elongation and, simultaneously, promote or inhibit the synthesis of gibberellin, that promote flowering.

Although growth retardants reduced ‘Lilliput’ internode length, main stem height, side branches and plant height (Table 1), plants were not short and compact enough to meet market quality standards. Considering the pot size used for ‘Lilliput’ production, a height pattern between 20-25 cm would be appropriated for selection of plants with a pleasant and compact aspect. Earlier application of retardants, when main stem internodes are

Table 3 - Effects of growth retardants on leaf mass ratio (LMR), foliage and inflorescences harvest index (FIHI), inflorescences harvest index (IHI), inflorescence diameter and production cycle of ‘Lilliput’.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Leaf mass ratio</th>
<th>Foliage and inflorescences harvest index</th>
<th>Inflorescences harvest index</th>
<th>Inflorescence diameter</th>
<th>Production cycle</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>g kg(^{-1})</td>
<td>cm</td>
<td>cm</td>
<td>days</td>
</tr>
<tr>
<td>Control</td>
<td>0.23 b</td>
<td>0.44 c</td>
<td>0.22 a</td>
<td>3.61 a</td>
<td>49.5 b</td>
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<td>0.26 ab</td>
<td>0.50 abc</td>
<td>0.25 a</td>
<td>3.58 a</td>
<td>53.1 ab</td>
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<tr>
<td>3.75 g L(^{-1}) daminozide</td>
<td>0.28 a</td>
<td>0.50 abc</td>
<td>0.22 a</td>
<td>3.55 a</td>
<td>53.1 ab</td>
</tr>
<tr>
<td>5.00 g L(^{-1}) daminozide</td>
<td>0.26 ab</td>
<td>0.51 ab</td>
<td>0.26 a</td>
<td>3.52 a</td>
<td>55.5 a</td>
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<tr>
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<td>0.25 ab</td>
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<td>0.75 mg a.i. paclobutrazol</td>
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<td>0.50 abc</td>
<td>0.24 a</td>
<td>3.53 a</td>
<td>51.8 ab</td>
</tr>
<tr>
<td>1.00 mg a.i. paclobutrazol</td>
<td>0.28 a</td>
<td>0.52 ab</td>
<td>0.25 a</td>
<td>3.47 a</td>
<td>50.1 b</td>
</tr>
<tr>
<td>1.00 g L(^{-1}) chlormequat</td>
<td>0.28 a</td>
<td>0.55 a</td>
<td>0.27 a</td>
<td>3.48 a</td>
<td>52.6 ab</td>
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<tr>
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<td>0.50</td>
<td>0.24</td>
<td>3.54</td>
<td>52.0</td>
</tr>
<tr>
<td>F test treatment</td>
<td>3.84**</td>
<td>4.54**</td>
<td>1.24**</td>
<td>0.35**</td>
<td>3.81*</td>
</tr>
<tr>
<td>HSD (Tukey's test)</td>
<td>0.04</td>
<td>0.07</td>
<td>0.08</td>
<td>0.41</td>
<td>4.80</td>
</tr>
<tr>
<td>C.V (%)</td>
<td>6.83</td>
<td>6.28</td>
<td>14.73</td>
<td>4.93</td>
<td>3.89</td>
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</tbody>
</table>

Mean separation within columns by Tukey’s multiple range test, 5% level; HSD = honestly significant difference; C.V = coefficient of variation; “ nonsignificant at \(P < 0.05\); ‘ significant difference at \(P < 0.05\); ” significant difference at \(P < 0.01\).

still shorter (plants with at least the third-leaf-pair completely expanded), could lead to a major control of elongation, not delaying floral transition. Under inductive flowering conditions, two-leaf-pair stage may correspond to the transition from vegetative to reproductive stage in zinnia (Kim et al., 1989). A second application of retardants may be necessary to help controlling of side branches elongation after reaching visible apical flower bud stage. This application may also contribute to the control of main stem internodes elongation up to the end of the production cycle, keeping retardants at growth inhibiting levels, since as reported by Sachs & Hackett (1972), there is a gradual decay of retardants at the active sites, by metabolism.

Further studies should adjust suitable concentrations of retardants to an earlier developmental stage application and to a second application at visible apical flower bud stage, considering the cumulative effect of more than one application without delaying production cycle or causing phytotoxic symptoms and excessive growth suppression resulting in over-dwarfed plants.

Figure 5 - Regression models of daminozide and paclobutrazol effects on leaf mass ratio (A-B), foliage and inflorescences harvest index (C-D), inflorescences harvest index (E) and production cycle (F) of ‘Lilliput’. Vertical bars represent the standart error of mean, n = 4; *significant difference at (P < 0.05); **significant difference at (P < 0.01).

CONCLUSIONS

Daminozide (2.5 and 3.75 g L⁻¹), paclobutrazol (0.5; 0.75 and 1.0 mg a.i. per pot) and chlormequat (1.0 g L⁻¹) suppressed ‘Lilliput’ growth and did not affect flower diameter and production cycle, but plants were not short enough to meet market quality standards.

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