Ectoparasitic fauna in freshwater ornamental fish acquired by a wholesaler in the city of São Paulo

Fauna ectoparasitária em peixes ornamentais de água doce adquiridos por estabelecimento distribuidor na cidade de São Paulo

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Abstract
This study analyzed the ectoparasitic fauna of freshwater ornamental fish marketed by a wholesaler in the São Paulo city. This study was undertaken from January to December 2016. A total of 268 freshwater ornamental fishes of various species that showed signs of disease after arrival at the establishment were evaluated. From the individuals, ectoparasites were collected by scraping off the skin and gills and visualized using an optical microscope at increasing magnification of 40x, 100x, 200x, and 400x. Parasites were found in 220 animals (82%); 11 different parasites were recorded, which included Ichthyophthirius multifiliis, Monogenes, Metacercariae (Centrocestus formosanus), Epistylis spp., Chilodonella spp., Lernaea spp., Tetrahymena spp., Piscinoodinium spp., Trichodina spp., Ichthyobodo spp. and Argulus spp. The occurrence of the parasites in the establishment studied can indicate that the suppliers’ lack of good sanitary and hygiene practices in the management of their facilities, consequently distributing infested animals and putting the whole supply chain at risk.

Keywords: Ornamental fish. Ectoparasitic fauna. Diseases. Animal health.

Introduction
The ornamental aquarium fish constitutes a large segment of the pet industry in which the United States, Europe, and Japan make up the bulk of the market (NOGA, 2010). Brazil holds the second place in the world market for pets. In 2016, it was valued at R$ 18.9 billion, according to the Brazilian Institute of Geography and Statistics. Fish are the fourth most common pet species in Brazil, behind dogs, cats and birds, and accounts for about 18 million units according to the latest survey (ABINPET, 2017).
Around the world, there are 4000 and 5000 ornamental freshwater fish species (of which about 90% are already cultivated) and over 1450 ornamental marine species, most of which are extracted from nature (WHITTINGTON; CHONG, 2007). Most ornamental fish traded in Brazil are cultivated on fish farms. In Brazil, the Minas Gerais state is the largest producer and supplies part of the market in the Northeast, Midwest, Southeast, and South of the country (MAGALHÃES; JACOBI, 2013). The Amazon state has the greatest diversity of ornamental fishes, with a predominance of extractivism. The Amazon state contributes about 93% to the Brazilian ornamental fish exports, which is an important economic activity that generates employment and income for a part of the population (ANJOS et al., 2009).

The trade and movement of live animals without standardized and safe technical procedures increases the risk of numerous agents that cause infectious diseases and serious health problems to spread, especially in poor regions (WHITTINGTON; CHONG, 2007; PEELER; FEIST, 2011; MCDERMOTT; PALMEIRO, 2013). Animals are often exposed to excessive handling and crowding in transport (DAVIS et al., 2002), which increases the concentration of metabolic residues (mainly ammonia) in water, leading to animal intoxication and stress. Stress has inhibitory effects on the immune system and favors the onset of opportunistic diseases such as some parasites (FRANCIS-FLOYD, 2012). Animals often live in harmony with the pathogenic agents, but as soon as an imbalance occurs, as in the immunity of the host, the latter proliferate, causing diseases (CARDOSO et al., 2017a, 2017b). This study aimed to evaluate animals that presented clinical signs compatible with ectoparasitic disease after they were transported to an ornamental fish wholesaler located in São Paulo city.

Materials and Methods

Study Animals

The place chosen for the study was a wholesale shop located in the São Paulo city that receive animals from different parts of Brazil to distribute to all Brazilian states. The volume of animals traded weekly is around fifty thousands of fresh and marine fishes.

Materials were collected from January to December 2016. The species chosen for the study were those with high history of mortality in the establishment, and the animals used were those with apathy and clinical signs of the disease. Skin scrapings and gills from 268 freshwater fishes of different species were analyzed.

Sample Collection

Moribund animals were treated with an anesthetic solution of Eugenol – 75 mg/L of clove oil (ROUBACH et al., 2005) diluted in a 4-liter container for 40 to 60 minutes and then euthanized by the spinal cord section technique (NOGA, 2010). Once euthanized, the fish skin and gills were scraped with a scalpel. This study was approved by Ethics Committee, under number CEUA N 4543160516.

Sample Analysis

For the analysis of the samples collected by scraping, a drop of water was deposited on the slide to facilitate the observation of the movement of the ectoparasites under the microscope cover slip (NOGA, 2010; MARTINS et al., 2015). The material was analyzed under an optical microscope at increasing magnifications of 40 ×, 100 ×, 200 ×, and 400 ×.

Results and Discussion

Among the 268 individuals examined under the microscope, 215 (80.22%) showed infection by ectoparasites. (Table 1).

<table>
<thead>
<tr>
<th>Agent</th>
<th>No. of fish infected</th>
<th>% of parasites</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Ichthyophthirius multifiliis</em></td>
<td>75</td>
<td>34.091</td>
</tr>
<tr>
<td>Metacercariae</td>
<td>40</td>
<td>18.182</td>
</tr>
<tr>
<td>Monogenea</td>
<td>38</td>
<td>17.273</td>
</tr>
<tr>
<td><em>Epistylis</em> spp.</td>
<td>24</td>
<td>10.909</td>
</tr>
<tr>
<td><em>Chilodonella</em> spp.</td>
<td>11</td>
<td>5.000</td>
</tr>
<tr>
<td><em>Lernaea</em> spp.</td>
<td>10</td>
<td>4.545</td>
</tr>
<tr>
<td>Tetrahymena spp.</td>
<td>6</td>
<td>2.727</td>
</tr>
<tr>
<td><em>Piscinoodinium</em> spp.</td>
<td>5</td>
<td>2.273</td>
</tr>
<tr>
<td><em>Trichodina</em> spp.</td>
<td>5</td>
<td>2.273</td>
</tr>
<tr>
<td><em>Ichthyobodo</em> spp.</td>
<td>4</td>
<td>1.818</td>
</tr>
<tr>
<td>Argulus spp.</td>
<td>2</td>
<td>0.909</td>
</tr>
</tbody>
</table>

TOTAL PARASITES 220

ANIMALS INFECTED 215 80.224

TOTAL ANIMALS 268

Table 1 – Parasites found in ornamental fish were evaluated. Only presence (1 parasite) or absence (0 parasites) was noted. The study was undertaken at the Department of Preventive Veterinary Medicine and Animal Health, Faculty of Veterinary Medicine and Animal Science, University of São Paulo – USP, São Paulo (SP) in 2017.
Ichthyophthirius multifiliis causes the white spot disease (Figure 1a). When analyzed, they present a horseshoe-shaped nucleus easily recognizable (Figure 1b). This parasite species causes the most trouble for hobbyists around the world (TAVARES-DIAS et al., 2010; IQBAL; HAROON, 2014; ADEL et al., 2015; SAHA; BANDYOPADHYAY, 2017). The disease is easily identified through clinical signs and microscopy of the scraping of the affected area. A single parasite gives rise to hundreds of others that are ready to infect new hosts, and consequently they spread quickly. Overcrowded aquariums are mainly affected. This parasite can lead to high mortality rates. When diagnosed early and treated, recovery is successful with reduced losses (FRANCIS-FLOYD et al., 2009; VON GERSDORFF JØRGENSEN, 2016).

![Figure 1](image1.png)

Figure 1 – (a) White spot disease, caused by the protozoan Ichthyophthirius multifiliis in the ornamental fish Amphilophus trimaculatus; (b) The mature parasite under 200 × magnification showing the horseshoe-shaped core – photo by CARDOSO, PEDRO HENRIQUE MAGALHÃES

Other protozoan ectoparasites prevalent at the study site include Ichthyobodo spp., Epistyli ssp., Chilodonella spp., Tetrahymena spp., Piscinoodinium spp., Trichodina spp., which are frequently reported from other studies across the world (THILAKARATNE et al., 2003; TAVARES-DIAS et al., 2010; IQBAL; HAROON, 2014; SAHA; BANDYOPADHYAY, 2017). They are almost frequently associated with aquatic environments with low water quality, high concentration of organic matter, and inefficient sanitary management. These parasites infest debilitated hosts, causing intense stress, serious injury, leading to death, with significant economic losses (MARTINS et al., 2015).

Many of the fishes analyzed (Figure 2a) presented metacercariae in their gills, (Figure 2b) suggestive of the parasite Centrocestus sp. The life cycle of this parasite requires a snail as a transport host. Good management practices that prevent molluscs from entering the system are sufficient to control them. Fishes with high infestations by encapsulated metacercariae in the gills presented respiratory deficiency and panting behavior (MEHRDANA et al., 2014). Among the fishes that had metacercariae, 38% were of the species Xiphophorus maculatus, whose farmer reported the presence of snails and other animals including birds, insects, amphibians, and mammals in the property, which implied a lack of sufficient sanitation at this facility.

![Figure 2](image2.png)

Figure 2 – (a) Xiphophorus maculatus, which was very weak and breathless; (b) The presence of metacercariae (shown at 40 × magnification) suggests Centrocestus sp. in the gills – photo by CARDOSO, PEDRO HENRIQUE MAGALHÃES

Species of the class Monogenea are also importance parasites affecting freshwater fishes. The most frequent genera are Dactylogyrus and Gyrodactylus. These parasites have hooks (Figure 3b) that attach to the skin tissues (Figure 3a) and gills of the host, causing lesions that open doors for secondary bacterial infections (REED et al., 2012; CARDOSO et al., 2017b). Other parasites found less frequently were Lernaea, Piscinoodinium, and Argulus. In addition to parasitizing fishes, the crustaceans Lernaea (Figures 4a and 4b) and Argulus (Figures 4c and 4d) also cause lesions that may serve as a gateway for secondary bacterial infections and disseminate viral diseases (STECKLER; YANONG, 2012a, 2012b). Piscinoodinium is a parasite without any specific host, which is responsible for significant sanitary problems in Brazil in freshwater fishes, according to producers. Intense infestations cause high mortality rates (MARTINS et al., 2015) if not treated early on.

![Figure 3](image3.png)

Figure 3 – (a) Polypterus palmas with Monogenes on the skin and ocular region; (b) parasite (shown at 40 × magnification) suggests Gyrodactylus spp. – photo by CARDOSO, PEDRO HENRIQUE MAGALHÃES

![Figure 4](image4.png)

Figure 4 – (a) Lernaea with 10× magnification; (b) Argulus with 40× magnification
Figure 4 – (a) *Lernaea* spp. (arrow) parasitizing *Xiphophorus hellerii*; (b) 40 × magnification of the anchor-shaped hooks that help to attach to the skin of the fish can be seen; (c) *Argulus* spp. parasitizing *Cyprinus carpio*; (d) *Argulus* spp. under 4 × magnification. – photo by CARDOSO, PEDRO HENRIQUE MAGALHÃES CARDOSO

The complete list of species examined is presented in Table 2.

**Table 2** – Species of fish evaluated in this study. This study was undertaken at the Department of Preventive Veterinary Medicine and Animal Health, Faculty of Veterinary Medicine and Animal Science, University of São Paulo – USP, São Paulo (SP) in 2017.

<table>
<thead>
<tr>
<th>Species</th>
<th>No. of animals examined</th>
<th>Species</th>
<th>No. of animals examined</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Aequidens rivulatus</em></td>
<td>3</td>
<td><em>Metynnis maculatus</em></td>
<td>1</td>
</tr>
<tr>
<td><em>Amphilophus citrinellus</em></td>
<td>1</td>
<td><em>Mikrogeophagus alitispinosus</em></td>
<td>7</td>
</tr>
<tr>
<td><em>Amphilophus trimaculatus</em></td>
<td>4</td>
<td><em>Mikrogeophagus ramirezi</em></td>
<td>4</td>
</tr>
<tr>
<td><em>Arapaima gigas</em></td>
<td>1</td>
<td><em>Misgurnus anguilicaudatus</em></td>
<td>1</td>
</tr>
<tr>
<td><em>Astronotus ocellatus</em></td>
<td>3</td>
<td><em>Moenkhausia sanctaehlenae</em></td>
<td>1</td>
</tr>
<tr>
<td><em>Aulonocara hansbaenschi</em></td>
<td>1</td>
<td><em>Monodactylus argenteus</em></td>
<td>1</td>
</tr>
<tr>
<td><em>Baryancistrus</em> ssp.</td>
<td>2</td>
<td><em>Myloplus rubripinnis</em></td>
<td>2</td>
</tr>
<tr>
<td><em>Betta splendens</em></td>
<td>6</td>
<td><em>Nandopsis octofasciatum</em></td>
<td>1</td>
</tr>
<tr>
<td><em>Carassius auratus</em></td>
<td>31</td>
<td><em>Nematobrycon palmeri</em></td>
<td>1</td>
</tr>
<tr>
<td><em>Carnegiella strigata</em></td>
<td>2</td>
<td><em>Notopterus chitala</em></td>
<td>1</td>
</tr>
<tr>
<td><em>Chromobotia macracanthus</em></td>
<td>2</td>
<td><em>Osteoglossum bicirrhosus</em></td>
<td>1</td>
</tr>
<tr>
<td><em>Cichla ocellaris</em></td>
<td>1</td>
<td><em>Otocinclus</em> spp.</td>
<td>1</td>
</tr>
</tbody>
</table>

**TOTAL** 268

Carassius auratus, Xiphophorus maculatus, Pterophyllum scalare, Trichodus spp., and Poecilia reticulata (together responsible for 40.30% of the animals examined) showed parasite infestation. However, additional studies with a higher number of fish of species with clinical signs are required to verify whether the disease is caused by parasitism or another infectious disease.

**Conclusion**

The occurrence of the aforementioned parasites in the establishment under study can indicate the suppliers’ lack...
of good sanitary and hygiene practices in the management of their facilities, consequently distributing infested animals and putting the whole supply chain at risk. This causes losses, with the death of animals and increased medical expenditure to treat infected animals.

These findings highlight the need for intensifying a system of good practices of health management at the suppliers’ facility, such as: identification of parasites at source and adequate measures to treat, control or avoid parasite outbreaks before sending fishes. This would be possible with the presence of a qualified veterinary professional in these facilities to provide technical guidance.

**Acknowledgements**

We thank the company Ipiranga Peixes Ornamentais from which we procured the ornamental fishes for this study for kindly donating the animals.

**Conflict of Interest**

There is no conflict of interest

**References**


