

EFFECT OF AN IODOPHOR IN CHILLING WATER ON THE BACTERIAL COUNTS OF PROCESSED POULTRY

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RFMV-A/17

PANETTA, J. C. & COELHO, W. P. *Effect of an iodophor in chilling water on the bacterial counts of processed poultry.* **Rev. Fac. Med. vet. Zootec. Univ. S. Paulo**, 13(1):249-52, 1976.

SUMMARY: *The experiment was conducted to determine whether or not the iodophor is effective in the decontamination of poultry carcasses immersed in the treated chilling water. 45 carcasses were used at weekly intervals, allocated in 9 groups of 5 each. 8 groups were treated by iodophor solutions in the chilling tank at the following dilutions: 1:250; 1:500; 1.750; 1:1000; 1:1250; 1:1500; 1:1750; 1:2000.*

One group remained as control. The samples for bacteriological examinations were collected by swabs. The use of iodophor in the water from the chilling tank resulted in the decontamination of the poultry carcasses at dilutions from 1:250 to 1:250. The experiments were repeated 10 times.

UNITERMS: *Poultry, carcasses *; Poultry, spoilate *; Water chilling, decontamination *.*

INTRODUCTION AND LITERATURE

A low bacteria count in the skin of processed carcasses represent good hygienic procedures. This improves the poultry products for marketing as well as increasing shelf life and bettering appearance of the poultry^{17, 18}.

These improvements in poultry processing have arisen from technologists' constant preoccupation in examination of the contamination of the skin in processed poultry and consideration of its importance with regards to the health of the public³.

Technological procedures have searched for minimising the incidence and development of the contaminants. Thus, with this objective in mind, the following methods were investigated: chlorination of the water⁵; the use of antibiotics¹; the use of sodium chloride solutions⁹; the utilisation of phosphate and poly-phosphate solutions⁶; the use of acid solutions¹⁵; the application of carbon dioxide atmosphere¹³; the use of new package¹²; shift of the pre-chilling procedures⁴; in the processing line¹⁴.

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As many authors^{8, 19} have identified the chilling tank as an important factor in the contamination of processed poultry, the following experiment was conducted to determine whether or not the iodophor acts in the decontamination of poultry carcasses when diluted in water for immersion chilling of poultry.

EXPERIMENTAL PROCEDURE

In this work the authors choose for bacteriological examinations the total count of aerobics (at 35°C), the count of coliform group (which has *E. coli* as pattern) and the count of enterococcus group (which has *S. faecalis* as pattern). This practice is recommended by several authors^{16, 20} who consider coliforms and enterococcus as fecal indicator organisms which show the hygienic quality of the processed carcasses.

Poultry carcasses from a commercial processing plant in S. Paulo City were used in the experiment.

At weekly intervals 45 carcasses were allocated, at random, in 9 groups of 5 each. 8 group were treated by iodophor* diluted in the chilling water, plus ice rocks, at the following dilutions: 1:250; 1:500; 1:750; 1:1000; 1:1250; 1:1500; 1:1750; 1:2000.

One group chilled by conventional procedure, remained as the control.

The treated poultry carcasses were immersed in the solutions of iodophor contained in the stainless steel chilling tank for 15 minutes, under continuous agitation. The carcasses were sent for packaging in plastic bags and transferred to the laboratory for bacteriological examinations.

The five carcasses from the control group were randomised after the packaging in plastic bags during the conventional processing for immersion chilling, in which non-treated water, plus ice rocks, was used in a stainless steel tank.

In the laboratory the samples are obtained by swabbing of the carcass skin, according to the technique written by WALKER & AYRES¹⁸, but with the changes

proposed by KOTULA¹⁰, that replaced the conventional metal instruments by sterilised cotton.

The samples were collected by swabs from the external areas of the carcass corresponding to 8 square centimeters (2 cm² under the left wing, 2 cm² under the right wing and 4 cm² in the breast).

For determination of the total numbers of coliforms, enterococcus and aerobics, the 3 swabs from each carcass were put in 250 ml, flasks with 100 ml, of Ringer's solution and 1 gram of peptone sterilised at 120°C, under 15 lbs pressure. After this, the samples were sent to the microbiological laboratory of The Veterinary School, for examination (transportation taking 20 minutes).

The following culture media were used in these experiments: "plate count agar" (Oxoid 37) for aerobics; "brilliant green bile broth" (Difco B-7) for coliforms; the medium from LITSKY, MALMANN & FIELD¹¹, modified, for enterococcus.

The bacterial computation was made according to the method adopted by PANETTA¹⁴ in previous work.

RESULTS AND DISCUSSION

The treatment of the chilling water by iodophor reduced the degree of bacterial contamination of the poultry carcasses. There was complete decontamination of the processed poultry carcasses by use of iodophor dilutions from 1:250 to 1:1250. The presence of bacteria in the surface of the poultry carcasses was observed in the 1:1500 dilution.

The Table 1 shows the counts of total aerobics (at 35°C) coliforms and enterococcus. These results represent the averages from the tests carried out, i.e., one test per week over a period of 10 weeks, each test involving 9 groups of 5 carcasses.

From the practical standpoint the 1:1250 dilution was chosen for the treatment of the water used for immersion chilling during the poultry processing.

* Biocid, Pfizer's trade name for an iodophor from Vanodine International Limited, England.

TABLE 1

Average counts (10 replications) of total aerobies (at 35°C), coliforms and enterococcus obtained in processed poultry carcasses treated and non-treated by iodophor.

Iodophor dilutions	N.º of bacteria per square centimeter (x)		
	Aerobies	Coliforms	Enterococcus
1: 250	—	—	—
1: 500	—	—	—
1: 750	—	—	—
1:1000	—	—	—
1:1250	—	—	—
1:1500	35.0 x 10 ²	7.0 x 10 ²	21.0 x 10 ²
1:1750	58.0 x 10 ²	12.0 x 10 ²	37.0 x 10 ²
1:2000	11.2 x 10 ³	34.0 x 10 ²	66.0 x 10 ²
Control	10.5 x 10 ⁴	28.3 x 10 ³	67.9 x 10 ³

Special attention should be paid to the higher enterococcus levels over coliforms. This is an extremely important fact for the bacteriological quality of the water from the chilling tank was taken into consideration when selecting a representative bacterium¹⁶. On this subject, the enterococcus as contaminant of the chilling

water will be studied in further work because from the experimental results it is evident that they appear to have more resistance to the temperature of the water used in the chilling tank. This point is endorsed by PANETTA¹¹ and WILKERSON²⁰.

RFMV-A/17

PANETTA, J. C. & COELHO, W. P. *Efeito de um "iodophor" na descontaminação de carcaças de frangos industrializados* **Rev. Fac. Med. vet. Zootec. Univ. S. Paulo**, 13(1):249-52, 1976.

RESUMO: O trabalho foi conduzido com o propósito de testar a eficiência de um "iodophor" na descontaminação da água do tanque de pré-resfriamento utilizado na industrialização de carcaças de aves. Foram usadas 45 carcaças de frangos, a intervalos semanais, distribuídas em 9 grupos de carcaças cada. 8 grupos foram tratados pelo "iodophor" adicionado à água do tanque de pré-resfriamento nas seguintes diluições: 1:250; 1:500; 1:750; 1:1000; 1:1250; 1:1500; 1:1750 e 1:2000. Um grupo foi reservado como controle. As amostras para o exame bacteriológico foram colhidas por "swab" da pele das carcaças. A utilização do iodophor diluído na água do tanque resultou na descontaminação das carcaças, quando as diluições localizaram-se entre 1:250 e 1:1250. O experimento foi replicado dez vezes.

UNITERMOS: Aves, carcaças*; Carne, deterioração*; Frangos, industrialização*.

REFERÊNCIAS BIBLIOGRÁFICAS

- 1 — AYRES, J.C.; WALKER, H.W.; FANELLI, M.J.; KING, A.W.; THOMAS, F. Use of antibiotics in prolonging storage life of dressed chicken. *Food Technol.*, **10**(11):563-68, 1956.
- 2 — BARNES, E.M. The effect of chlorinating chill tanks of the bacteriological condition of processed chickens. Institute Internationale du Froid. Bulletin-Commission 4-Karlsruhe — 1-7, 1965.
- 3 — BARNES, E.M. & SHRIMPTON, D.H. The effect of processing and marketing procedure on the bacteriological condition and shelf life of eviscerated turkeys. *Brit. Poult. Sci.*, **9**(3):243-51, 1968.
- 4 — CASALE, J.O.; MAY, K.N.; POWERS, J.J. Effects of three chilling methods on bacteriological organoleptic and physical properties of broiler chickens. *Food Technol.*, **19**: 859-61, 1965.
- 5 — DAWSON, L.E.; MALLMANN, W.L.; FRANG, M.; WALTERS, S. The influence of chlorine treatments on bacterial population and taste panel evaluation of chicken fryers. *Poultry Sci.*, **35**:1140, 1956.
- 6 — ELLIOT, R.P.; STRAKA, R.P.; GARBALDI, J.A. Effect of polyphosphates on growth of pseudomonas from poultry meat. *Bacteriol Proceedings Amer. Soc. Microbiol.*, **1**, 1964a.
- 7 — FROMM, D. Influence of re-using chill tank slush ice on market quality of eviscerated broilers. *Food Technol.*, **12**(5):257-59, 1968.
- 8 — GALTON, M.M. et al. Salmonellosis in poultry processing plants in Florida. *Amer. J. Vet. Res.*, **10**(58): 132-137, 1955.
- 9 — GARDNER, F.A. & ATKINSON, R.L. Tissue changes associated with chilling broilers in sodium chloride solutions. *Poultry Sci.*, **46**:1262, 1967.
- 10 — KOTULA, A.W. Variability in microbiological samplings of chickens by the swab method. *Poultry Sci.*, **45** (2):233-236, 1966.
- 11 — LITSKY, W. et al. A new medium for the detections of enterococci in water. *Amer. J. publ. Hlth.*, **43**(7): 873-79, 1953.
- 12 — MAY, K.N.; POWELL, W.R.; HUDSPETH, J.P. A comparison of quality of fresh chicken packed in various containers. Georgia Ag. Expt. Sta. Bulletin N.S. 168, 1966.
- 13 — OGILVY, W.S. & AYRES, J.C. Post-mortem changes in stored meats. II. The effect of atmosphere containing carbon dioxide in prolonging the storage life of cut-up. *Food Technol.*, **5**:97-102, 1951.
- 14 — PANETTA, J.C. Determinação de alguns contaminantes de frangos abatidos num matadouro de São Paulo e seu comportamento em face de modificações introduzidas na linha industrial. *Rev. Fac. Med. vet. Zootec. Univ. S. Paulo*, **9**:73-92, 1972.
- 15 — PERRY, G.A.; LAWRENCE, R.L.; MELNICK, D. Extension of poultry shelf life by processing with sorbic acid. *Food. Technol.*, **18**:891-97, 1964.
- 16 — RAJ, H. Detection and enumeration of fecal indicator organisms in frozen sea foods. II. Enterococci. *Appl. Microbiol.*, **9**(4):295-303, 1961.
- 17 — SURKIEWICZ, B.F.; JOHNSTON, R.W.; MORAN, A.B.; KRUMM, G.W. A bacteriological survey of chicken eviscerating plants. *Food Technol.*, **23**:1066-1069, 1969.
- 18 — WALKER, H.W. & AYRES, J.C. Incidence and kinds of microorganisms associated with commercially dressed poultry. *Applied Microbiol.*, **4**:345-49, 1956.
- 19 — WILDER, A.N. & MACREADY, R.A. Isolation of Salmonella from poultry. *New Engl. J. Med.*, **274**(26): 1453-60, 1966.
- 20 — WILKERSON, W.B. et al. Occurrence of enterococci and coliform organisms on fresh and stored poultry. *Food Technol.*, **15**(6):286-92, 1961.

Recebido para publicação em 15-3-76
Aprovado para publicação em 25-3-76